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HAL Id: insu-00371767
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Submitted on 1 Apr 2009

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Raman quantification factor calibration for CO-CO$_2$ gas mixture in synthetic fluid inclusions: Application to oxygen fugacity calculation in magmatic systems.

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Abstract:

With a combined approach using Solid State $^{13}$C-MAS NMR and Laser Raman Microspectroscopy, we investigated the CO$_2$-CO gas composition ($X$(CO$_2$)) in fluid inclusions synthesised under high pressure (200-300 MPa), high temperature (1225-1250°C) and reducing conditions ($17 < P$(H$_2$) $< 62$ bars). Fluid inclusions are entrapped in a volatile-bearing basaltic glass which was characterized by FTIR for determining the water solubility (H$_2$O$^m$).

$^{13}$C-MAS NMR is used as a standard analysis for determining the $X$(CO$_2$). The Raman quantification factors between $^{13}$CO$_2$ and $^{13}$CO is determined from peak area (F-factor), peak height (G-factor) and according to the Placzek’s polarizability theory. The calibration is derived for both CO$_2$ Fermi diad resonances: 2$\nu_2$ and $\nu_1$.

We obtain similar values for the main CO$_2$ resonance (2$\nu_2$) with 1.956 and 1.809 for F and G respectively. Results are consistent with the fact that peak height and area will measure the same quantity. For $\nu_1$, multiple calibration trends are observed. The different trends are explained by the different $^{13}$C/$^{12}$C ratio observed in between the samples. However, we suggest that such resonance is not suitable for determining the fluid inclusion compositions.

We extended the $^{13}$C results for calibrating the F- and G-factors for $^{12}$CO$_2$-$^{12}$CO gas mixture in the fluid inclusions and for the main CO$_2$ resonance. For $^{12}$CO$_2$-$^{12}$CO mixture, F and G values are 1.856 and 1.756 which is in the same order as the derived values for $^{13}$C species. Thus, we propose that no significant $^{13}$C/$^{12}$C fractionation occurs in the fluid phase and both isotopes will behave in a similar way.
Using the derived calibration for $^{12}\text{C}$ and $^{13}\text{C}$ species, the $X(\text{CO}_2)$ in the fluid phase was recalculated. Results are similar for both isotopes witnessing the similar behaviour of $^{12}\text{C}$ and $^{13}\text{C}$ fluid species during the experiments.

The log $f(O_2)$ experienced by each sample has been calculated through a thermodynamic approach using 2 independent methods. The log $f(O_2)$ calculated from the $\text{H}_2\text{O}^{\text{m}}$ in the glass and the $X(\text{CO}_2)$ in the fluid phase are in good agreement. Large discrepancy is observed for low $\text{H}_2\text{O}^{\text{m}}$ content which gives lower log $f(O_2)$ value than expected from experimental conditions. Large uncertainties on the $\text{H}_2\text{O}^{\text{m}}$ measurements will induce a very approximate value for the log $f(O_2)$. This method may not be accurate enough at low $\text{H}_2\text{O}^{\text{m}}$ and using the $X(\text{CO}_2)$ in the fluid phase would therefore provide a better estimate of the log $f(O_2)$.

**Keywords:** Fluid inclusion, Raman spectroscopy, Solid State NMR, Carbon dioxide, oxygen fugacity.
1 Introduction:

Volatile s are ubiquitous components implied in many geological systems. In particular, CO₂ and H₂O are the two most abundant volatiles emitted at volcanic centres (Gerlach and Graeber, 1985; Symonds et al., 1994; Jambon, 1994; Morizet et al., 2002). CO₂ is usually present in magmas with moderately oxidised conditions. However, under more reducing conditions other volatiles become stable such as CO, CH₄ (Kadik et al., 2004). Although the distinction should be made between volatiles dissolved in the melt and volatiles implied in the fluid phase in equilibrium with the melt, both need to be considered as parts of a given system. The solubilities of H₂O and CO₂ are controlled by H₂O and CO₂ fluid fugacities (Dixon et al. 1995; Dixon, 1997) for a given melt composition at fixed P and T.

In magmatic systems, upon decompression volatiles exsolve the melt phase to be stored in the fluid phase. This observation is even more verified for CO₂ which has a low solubility compared to H₂O (e.g., Blank and Brooker, 1994; Dixon, 1997; Morizet et al., 2002). The fluid phase is partly preserved as inclusions within minerals or within the silicate melt phase itself. Fluid inclusions study can provide valuable information on the Earth interiors volatiles content and degassing (Frezzotti and Peccerillo, 2007).

Estimating or calculating the oxygen fugacity (f(O₂)) in Earth materials is a recurring problem. The f(O₂) cannot be measured directly except in volcanic gases (Symonds et al., 1994). In the melt phase, the f(O₂) may be estimated using several methods. Within the melt, the Fe³⁺/Fe²⁺ is a proxy for the f(O₂) as described by Kress and Carmichael (1991) or Bezos and Humler (2005). The H₂O content in the melt depends on the f(H₂O) in the fluid which is in turn a function of the f(O₂) according to the reaction:
The $f$(H$_2$O) is first estimated from existing models (e.g. Dixon et al., 1995). Then, with a known $f$(H$_2$), one can estimate the $f$(O$_2$). The $f$(O$_2$) can also be calculated from the equilibrium of carbon-bearing volatile species and according to several reaction such as:

\[ H_2^\beta + \frac{1}{2}O_2^\beta \leftrightarrow H_2O^\beta \]  

(1)

\[ CO_2^\beta \leftrightarrow CO^\beta + \frac{1}{2}O_2^\beta \]  

(2)

\[ CO_2^\beta + 2H_2^\beta \leftrightarrow CH_4^\beta + O_2^\beta \]  

(3)

Either reaction can be used to calculate $f$(O$_2$) depending on the species abundances. The first reaction is commonly used in moderately reducing conditions, such as with present day MORB (Pawley et al. 1992). CO$_2$-rich fluid inclusions are also very common in shallow mantle rocks (Andersen and Neumann, 2001).

The second reaction occurs with more severe reducing conditions (Holloway, 1987; Kadik et al., 2004) where CH$_4$ is the dominant species and is supposed to have been a major control of the degassing during the Archean period (Arculus, 1985). CH$_4$ is identified in the fluid phase entrapped in inclusions for particular geological settings. Beeskow et al. (2005) analysed CH$_4$-CO$_2$ fluid inclusions in quartz collected from a coalfield in South Wales. Liu and Fei (2006) identified CH$_4$ in olivine-hosted fluid inclusions from Paleozoic ophiolites.

The investigation on the composition of the fluid phase is of great interest for the understanding of the Earth degassing. It thus appears compulsory to study the composition of the fluid phase as it provides valuable information on the redox state of the Earth mantle.
1.1 Fluid inclusions analyses: Quantification of the fluid species.

Qualitative and quantitative analyses of fluid inclusions can be achieved with several techniques either destructive or non-destructive. The different approaches are reviewed elsewhere (Dubessy et al., 1989; Roedder, 1990; Boiron and Dubessy, 1994; Shepherd et al., 1998).

Laser Raman Microspectrometry (LRM) has been widely used for fluid inclusion analyses as it is non-destructive and not time consuming (Roedder, 1990; Burke 2001). On the other hand, Raman is virtually non-quantitative (McMillan, 1984) and can only be used with relevant calibration with standards with known composition (Wopenka and Pasteris, 1987; Dubessy et al., 1989; Seitz et al., 1993). LRM Non-quantitative aspects arise from the lack of knowledge on the analysed sample and therefore the number of excited molecule. Thus only the mole ratios between species are usually used.

In fluid inclusion composition analyses, LRM technique is coupled with other techniques such as microthermometry (e.g. Bergman and Dubessy, 1984; Chen et al., 2004), FTIR (Wopenka et al., 1990), LA-ICP-MS (e.g. Shepherd et al., 1998; Stalder et al., 1998; Günther et al., 1998) or even NMR (Dereppe et al., 1994).

NMR is the only bulk non-destructive method. It can be used to investigate various elements: $^{23}$Na, $^{29}$Si, $^{27}$Al or/and $^{13}$C in solid state (Kohn et al. 1991; Neuville et al., 2006; Lee and Stebbins, 2003) but also $^{35}$Cl in fluid inclusions (Sherriff et al., 1987). Although it appears to be a powerful technique, it has sample requirements: 1) Fe-free composition, 2) relatively large sample (> 100 mg) and 3) possible isotopic enrichment of elements with nuclear spin.
To be quantitative LRM requires knowledge of the spectroscopic behaviour of the molecular species as a function of the molecular environment. Combining LRM with MAS NMR is one adequate solution for achieving qualitative and quantitative analyses of fluid inclusions.

The gas species molar fraction (X) in fluid inclusion can be determined according to the following equation:

$$X_a = \frac{A_a}{\sum \sigma_i \zeta_i} = \frac{A_a}{\sum \frac{A_i}{F_i}} = \frac{H_a}{\sum \frac{H_i}{G_i}}$$  \hspace{1cm} (4)

where $X_a$ is the molar fraction of the species $a$, $A$ stands for the area of the species observed by LRM, $\sigma$ is the Raman cross-section coefficient of a given vibrational Raman active mode of a specified molecule and $\zeta$ the Raman spectrometer instrumental efficiency for the Raman active mode of the specified molecule and at a given excitation wavelength. $\zeta$ is specific to one spectrometer and may be totally different for another spectrometer. Wopenka and Pasteris (1987) combined these two parameters and defined $F$ as the integrated Raman quantification factor. Similarly, a linear Raman quantification factor $G$ can be defined and related to the absolute height of the peak (H).

$\sigma$ is wavelength dependent (Dubessy et al., 1989; Burke, 2001). $\zeta$ is a parameter specific to each Raman apparatus and therefore needs to be determined for each Raman spectrometer (Kerkhof and Kisch, 1993). As a result, if one needs to perform LRM quantitative analyses of fluid inclusions, it is imperative to calibrate those parameters. Many studies (e.g. Wopenka and Pasteris, 1987; Kherkhof, 1988a; Chou et al., 1990)
suggested acquiring such empirical calibration through the use of standards with known compositions.

In our study we combined two methods (NMR and LRM) to calibrate the Raman quantification factor, $F$, for $^{12-13}$CO$_2$ and $^{12-13}$CO. One major advantage is the fully quantitative aspect of the NMR (the intensity of the signal is directly proportional to the molar fraction of the identified species). As a bulk method, NMR will averaged out the compositional heterogeneity which can observed by LRM from one fluid inclusion to another.

The review by Burke (2001) lists the reported values for various gas species $\sigma$. Currently, there is a lack of knowledge concerning the calibration of $^{13}$C species. For the $2\nu_2$ $^{13}$CO$_2$ vibration (overtone of the bending vibration $\nu_2$ for a CO$_2$ molecule), a value of 1.5 for $\sigma$ at 514 nm has been used by several authors (Rosasco et al., 1975; Dhamelincourt et al., 1979). Moreover, little is known about the behaviour of gases at high pressure (i.e. most of the F and $\sigma$ are obtained for low pressure confined inclusions). Chou et al. (1990) and Seitz et al. (1993, 1996) investigated the effect of pressure on the F and G factor (ratio between the F or G of the 2 species) for CH$_4$-N$_2$ and CH$_4$-CO$_2$ synthetic fluid inclusions mixtures. In both systems, a strong change in the F ratio is observed as a function of pressure up to 700 bars. However, no investigation has been performed at very high pressure (few hundreds MPa).

1.2 Research objectives: Quantitative analysis through LMR.

We have conducted an analytical study based on the combined use of LRM and $^{13}$C-MAS NMR for a series of synthetic CO$_2$-CO bearing fluid inclusions.
The objective was to obtain a reliable calibration for the analysis of CO₂-CO bearing fluid inclusions by LMR, in order to derive F and G ratios for the CO-CO₂ mixture. Results are valid for ¹³C isotopes but we tried to extend the results to ¹²C species. Results will be discussed. Comparison with previous published data is difficult as the derived calibration is only valid for our LRM equipment (LabRam Jobin-Yvon). Application and discussion on the oxygen fugacity conditions during the experiments is also addressed. The \( f(O_2) \) calculated using LRM fluid inclusion analyses is compared to the \( f(O_2) \) value calculated through the H₂O dissolved in the glass. The relatively good agreement between the two methods suggests the reliability of our calibration.

2 Experimental and Analytical methods:

2.1 Experimental synthesis:
Synthetic Fe-free basaltic glasses in equilibrium with excess COH fluid phase were synthesized under high pressure high temperature using an Internally Heated Pressure Vessel (IHPV). The starting composition was prepared from a mixture of oxides (SiO₂, Al₂O₃ and MgO) and carbonates (Na₂CO₃ and CaCO₃). A two stage preparation was employed: 1) SiO₂, Al₂O₃, MgO and CaCO₃ were mixed together and held at 1050°C for few hours in order to dissociate the calcium carbonate (the resulting powder should be then CO₂-free); and 2) the Na₂CO₃ is added to complete the composition and is the source of CO₂ during the experiment. The composition in wt.% is 54.8, 21.1, 8.1, 12.9 and 3.2 for SiO₂, Al₂O₃, MgO, CaO and Na₂O respectively. The CO₂ added to the composition through the sodium carbonate is at
1 wt.% level. The sodium carbonate source added is 100% $^{13}$C-doped which is the only C species with a nuclear spin to be analysed by Solid-State MAS NMR.

Small amount of H$_2$O (between 0 and 1 wt.%) is added at the bottom of the platinum capsules. Starting composition is loaded on top of the water and the platinum capsule is sealed. Experiments are conducted in IHPV with run duration between 2 to 3 hours to ensure equilibrium is reached. The experimental conditions are 200 and 300 MPa for pressure, 1225 and 1250°C for temperature. The details of the experimental conditions for each sample are reported in Table 1.

Under those P,T conditions, the 1 wt.% $^{13}$CO$_2$ added to the starting composition is well above the solubility saturation level for basaltic compositions (Fine and Stolper, 1986; Pan et al., 1991; Dixon et al., 1995). Therefore, we expect the melt to be in equilibrium with an excess fluid phase in which C-species dominate.

The procedure to achieve the experimental conditions is described elsewhere (e.g., Scaillet et al., 1992) and can be summarized as follow. The IHPV is loaded at room temperature with a known partial pressure of pure H$_2$ (P(H$_2$), see Table 1), Argon gas is then loaded up to roughly half the desired total pressure (P$_{tot}$) value (either 100 or 150 MPa). The temperature is then brought to the final value. Previous experiments have shown that under those P-T conditions the final estimated P(H$_2$) is therefore equivalent to roughly two times the initial loaded P(H$_2$). The final P(H$_2$) experienced during the run are shown in Table 1. At run conditions, the hydrogen diffuses into the Pt capsule and acts as reducing agent for the loaded CO$_2$ according to the reaction in the fluid phase:

$$CO_2^\alpha + H_2^\beta \leftrightarrow CO^\beta + H_2O^\alpha$$

(5)
where $fl$ stands for the gas in the fluid phase and $m$ stands for the species in the melt phase. Further reduction will produce CH$_4$ as shown by Eq. (3).

The water produced during those reactions will mostly enter into the melt ($H_2O^m$) as the solubility level is very high compared to CO$_2$ and little water will be observed into the fluid inclusions (see Table 2). The range of P(H$_2$) was chosen to 1) avoid graphite saturation and 2) minimize the presence of CH$_4$ (Eq. (3)). Although we mainly obtained a mixture of CO$_2$-CO in the fluid phase, CH$_4$ was still identified. No graphite was observed in the whole sample (glass and fluid inclusions as revealed by NMR analysis).

The sample is quenched to the ambient temperature by dropping the capsules into the cold part of the IHPV. During the quench, the fluid phase is mainly located at the top of the capsules; however a non-negligible part of the fluid phase will be entrapped as inclusions inside the glass.

Several chips from the recovered glasses were selected for micro-Raman and FTIR analyses. Remaining of the glass sample was then crushed for $^{13}$C-MAS NMR analysis.

2.2 Confocal Micro-Raman:

LRM was used to characterize the fluid species inside the inclusions. The Raman system is a Jobin-Yvon Labram spectrometer (focal distance = 300 mm) equipped with a 1800 gratings/mm CCD detector. The light source is an Ar laser Innova 300-5W from Coherent© operating at 514.5 nm. The analyses were performed in confocal mode (hole = 500 μm, slit = 200 μm) so the contribution of the surrounding glass is limited and the inclusion contributes to the majority of the Raman scattering signal.
Spectrometer was set in the region of the CO$_2$-CO scattering between 1100 and 2200 cm$^{-1}$. Analyses were also done in the CH$_4$ region (2900 cm$^{-1}$) to check its presence. The spectral frequency position was measured with the emission lines of Ne- and Hg-lamps. The accuracy stays within ± 1 cm$^{-1}$.

We used a variable output laser power between 25 and 200 mW. The acquisition time on a given inclusion was typically between 10 and 120 sec. We have tested the influence of laser power and acquisition time on two inclusions in KB16. No change in the ratio of the different species was observed with varying analysis conditions. The determined ratio standard deviation does not exceed 3% in value. All analytical conditions are reported in Appendix.

More than 10 different inclusions for KB3, 5, 8 and 16 were analyzed for the calibration of the Raman quantification factors.

For some inclusions (see Appendix), the LRM spectrum was acquired in oversaturation conditions of the detector for the main CO$_2$ peak. For example, in KB5-14 the $2\nu_2^{12}$CO$_2$ resonance is not measured as it was above the saturation level of the CCD detector. The aim was to increase the intensity of the minor constituent peak, the $2\nu_2^{13}$CO$_2$ resonance.

For KB10 and 12, we analyzed 2 inclusions. Those samples were not included into the calibration. They are used as an independent verification for the reliability of our work in the f(O$_2$) calculation.

2.3 Micro-FTIR analyses for glass water content:

Doubly polished plates were prepared from selected glass sections for micro-FTIR analyses in order to quantify the water content (H$_2$O$^m$). Near-IR spectra (between 4000
and 6000 cm\(^{-1}\)) were obtained for 100 \(\mu\)m spot size using a Continuum microscope attached to a Nicolet 5700 Fourier transform infrared spectrometer. A KBr beamsplitter and MCT-B detector were used to collect 128 scans at a resolution of 4 cm\(^{-1}\). The atmospheric contribution (in particular water) was reduced by enclosing the sample in a box purged with dry air for at least 20 minutes. Prior to sample acquisition, an instrumental background was subtracted. Several spectra were taken on different location in the glass plate. A volatile-free FTIR spectrum was taken on a dry glass synthesized at 1300°C, 1 atm. and was subtracted to each volatile-bearing samples spectrum.

2.4 \(^{13}\)C-MAS NMR:

\(^{13}\)C-MAS NMR spectra of the partially crushed samples were acquired on a 500 MHz Bruker Avance (11.7 T). A 4 mm CP/MAS “double bearing” probe was employed and the samples loaded in a ZrO\(_2\) rotor with Teflon end-cap. Rotors were spun at 10 kHz. The measurements were made at the \(^{13}\)C frequency (125.75 MHz). All spectra are referenced against TMS (Tetramethylsilane).

For all samples, \(^{13}\)C-MAS NMR spectra were acquired with single \(\pi/4\) excitation of 2 \(\mu\)s (SPE spectra). For both KB8 and 16, 13C-MAS NMR spectra were also acquired by using a rotor-synchronised spin-echo sequence \((\pi/2 - \tau - \pi - \tau - \text{acq.})\) with a \(\pi/2\) pulse of 4 \(\mu\)s and \(\tau\) equal to one rotor period. In all cases, \(^1\)H decoupling during acquisition (SPINAL64 scheme with a rf field of 60 kHz) was used to prevent from broadening of the \(^{13}\)C lines due to heteronuclear interactions with protons (Laws et al., 2002). However, due to the very sharp signature of CO\(_2\) and CO in the fluid phase and the presence of
protons only in the surrounding glass, the broadening effect is unlikely to occur. During the evolution time in the spin-echo sequence ($\tau$), both CO$_2$ and CO may evolve differently; however it is unlikely it will produce a large difference on the quantification for this gaseous species especially for 1 rotor period (0.1 msec.) where the evolution is relatively small.

C-species can have long spin lattice relaxation times in silicate glasses (several minutes, Kohn et al., 1991). It is not the case in fluid phases where relaxation timescale are very short, we used several recycle time D1 (2, 20 sec., see Table 1 and Figure 2A) values to check the consistency in the response of the C-species.

The number of scans collected is relatively large because of the low CO$_2$ solubility in the melt and the small quantity of fluid inclusions (the most important part of the fluid escapes during the opening of the capsule). Thus, depending on D1 values, between 3k and 103k scans have been co-added.

3 Results:

3.1 Description of fluid inclusions:

Figure 1 shows typical fluid inclusions observed within the samples. Inclusions are disseminated but can be locally concentrated. The inclusions are entrapped during the quench in a glass matrix either clear in the case of KB16, KB8, KB10 and KB12 or cloudy in KB3 and 5. The cloudiness is due to the presence of crystals in the glass. The nature of the crystal is not fully known but we suggest it is unreacted material preserved during the quench. Due to the peraluminous character of the starting composition (21 wt.% Al$_2$O$_3$), we suppose the unreacted material are alumina or mullite crystals.
The inclusions are round-shaped and therefore considered as primary (Roedder, 1984) with sizes ranging from ~10 to 100 μm. Larger inclusions were seen but are very often pierced. The inclusions are clear and do not present either graphite lining or solid independent phase inside (Roedder, 1984).

3.2 Fluid inclusion spectrum analyses:

3.2.1 13C-MAS NMR spectra:

Figure 2A shows NMR spectra for KB3 (acquired with different recycle delay, D1 = 2 and 20 sec.) and KB5. Two major peaks are observed on the spectra: one located at 125 ppm and assigned to $^{13}$CO$_2$ and one located at 183.5 ppm and assigned to $^{13}$CO (Kohn et al., 1991). No peak is observed at around 0 ppm which would reveal the presence of $^{13}$CH$_4$ (Ripmeester and Ratcliffe, 1988; Subramanian et al., 2000). The same observation is made for the other samples. There is a weak broad peak visible in KB5 between 160 and 175 ppm which is attributed to carbonate groups (CO$_3^{2-}$). CO$_2$ dissolves in basaltic melt only as CO$_3^{2-}$ units (Blank and Brooker, 1994). The presence of such a broad signal is given by the molecular environment of CO$_3^{2-}$ dissolved in the glass. On the other hand, the presence of CO$_2$ and CO can only be assumed to be entrapped into fluid inclusions and not dissolved in the glass.

Experiments were performed in water undersaturated conditions (i.e. most of the H$_2$O and H$_2$ will dissolve in the glass, Gaillard et al., 2003). If H is present, it will reduce CO$_2$ to form CH$_4$. 
LRM suggests the presence of CH$_4$ in the fluid inclusion. However, the CH$_4$ concentration is estimated to be very small and therefore below the NMR detection limit (a few %).

The CO$_2$ molar fraction (X($^{13}$CO$_2$)) is calculated using the ratio of the integrated area underneath the peaks at 125 and 180 ppm.

Using several D1 time does not modify the ratio of the C species (Table 1 and Figure 2A). In KB3, for D1 = 2 sec., the X($^{13}$CO$_2$) is equal to 0.728. X(CO$_2$) for D1 = 20 sec. differs slightly and is equal to 0.770. The difference is probably within error and the standard deviation (0.029) between the 2 values will be used for the other samples measurements of X(CO$_2$). In Table 1, we report 0.749 as the average of the 2 values. The same applies for the 2 NMR analytical conditions used for KB16 (SPE and spin-echo spectra). Only a slight difference is observed in between the X($^{13}$CO$_2$) values from 0.689 for SPE to 0.634 for spin-echo spectrum. The error bar reported for KB16 (± 0.038, Table 1) is probably the maximum error that we can observe. This value will be used later in the error propagation calculation for correcting the data by water activity. It should be mentioned that those values are not corrected for H$_2$O molar fraction (X(H$_2$O)) in the fluid phase (see section 4.3).

3.2.2 Confocal micro-Raman spectra:

Typical Raman spectra are reported in Figure 2B for several samples. The dashed line underneath corresponds to a typical baseline placed prior to the integration measurements.
Several peaks are observed. Two groups of peaks are located: 1) in the 1300 cm\(^{-1}\) region and is assigned to CO\(_2\) molecules and 2) in 2100 cm\(^{-1}\) region and corresponds to CO molecules. However within each group several peaks are seen. For the CO molecules, 2 peaks are located at 2090 and 2137 cm\(^{-1}\) and are assigned to \(^{13}\)CO and \(^{12}\)CO respectively. The CO\(_2\) in the fluid phase is represented by the Fermi diad (Wienecke et al., 1986) at \(~1283, 1386\) cm\(^{-1}\) for \(^{12}\)CO\(_2\) and 1369, 1262 cm\(^{-1}\) for \(^{13}\)CO\(_2\). Each individual resonance corresponds to the \(2\nu_2\) and \(\nu_1\) respectively (Rosasco et al., 1975; Garrabos, 1980; Kerkhof and Olsen, 1990).

Additionally, we identified in KB16 (also identified in KB10 and 8) the CH\(_4\) symmetric stretch (\(\nu_1\)) at very low Raman shift value at 2900 cm\(^{-1}\) (usually \(~2915\) cm\(^{-1}\) as mentioned in Kerkhof, 1987; and Larsen et al., 1992). Because our samples are enriched in \(^{13}\)C isotope, we suggest this peak to be attributed to \(^{13}\)CH\(_4\). The intensity of the CH\(_4\) peak is lower than CO peak and we regard this species as a minor diluted component. For KB12 (sample with the lowest P(H\(_2\))), CH\(_4\) is totally absent from the spectrum or below detection limit. The small proportion of CH\(_4\) observed by LRM is in agreement with the NMR results which do not identify its presence.

The hot band near the Fermi diad of CO\(_2\) are the result of two effects (Rosso and Bodnar, 1995; Dubessy et al., 1999; Chen et al., 2004): 1) the anharmonicity of the vibrational levels producing different energetic differences between two successive vibrational levels; and 2) the population of the excited vibrational levels increases with temperature. In Figure 2B, only one hot band is visible and observed at 1406 cm\(^{-1}\). Other hot bands may be observed as we increase the temperature above 200°C.
Samples were supposed to exhibit 100% in $^{13}$C, but the incomplete decarbonatation of Ca$^{12}$CO$_3$ in the starting powder suggests $^{12}$C is still abundant and present as $^{12}$CO$_2$, $^{12}$CO in the fluid inclusions.

The signature of each species was quantified either by measuring the height (H) or the area (A) of the peaks. A typical baseline is represented in Figure 2B (dashed line) underneath the Raman spectrum.

Results from the integrated areas and heights of the different peaks (CO$_2$ 2$\nu_2$, $\nu_1$ and CO for $^{13}$C and $^{12}$C) are reported in the Appendix for the different analyzed inclusions. Peak areas or height have been measured with the built-in functions in Origin© software. A and H are proportional 1) to the quantity of analyzed molecules (function of fluid density, composition, size and depth of the inclusion), 2) to the Raman analytical conditions. For a given inclusion, increasing the acquisition time will increase the precision of the measurement.

From the integrated areas and the peak heights of the different species within the fluid inclusions, we can therefore calculate 1) the enrichment in $^{13}$C in each sample, 2) the fractionation $^{13}$C/$^{12}$C if it exists, 3) calibrate Raman quantification factor (F and G) by combining the NMR and the Raman results for $^{13}$C-species and thus 4) give an estimate of the molar fraction X($^{13}$CO$_2$) of each individual fluid inclusion which is used to calculate the $f$(O$_2$) the samples experienced during the run in equilibrium with the fluid phase. We extended further our calibration method to $^{12}$C-species. Raman quantification factor (F and G) and the X($^{12}$CO$_2$) are determined for $^{12}$C-species by combining the $^{13}$C-NMR results and the $^{12}$C-species Raman results.
3.2.3 Water content in the glass matrix (H\textsubscript{2}O\textsuperscript{m}): FTIR analyses.

The water content in the glass (H\textsubscript{2}O\textsuperscript{m}) was determined by micro-FTIR measurements. Quantification of the H\textsubscript{2}O\textsuperscript{m} is then used to estimate the X(H\textsubscript{2}O)\textsuperscript{f} as well as the f(O\textsubscript{2}) (see section 4).

In Figure 3, we show the FTIR spectra obtained for the basaltic glass in the water vibration region (KB10 is not shown for clarity). Two main water species are identified. One vibration located at 4500 cm\textsuperscript{-1} is assigned to OH groups and one at 5200 cm\textsuperscript{-1} is assigned to the bending of H\textsubscript{2}O in its molecular form (H\textsubscript{2}O\textsuperscript{mol}), consistent with previous studies (e.g., Stolper, 1982a; Ihinger et al., 1994; King et al., 2002; Stuke et al., 2006). The H\textsubscript{2}O\textsuperscript{mol} species are observed for samples with the highest water content whereas OH groups are always present which is in agreement with H\textsubscript{2}O speciation models.

The H\textsubscript{2}O\textsuperscript{m} is determined by quantifying the sum of each individual species (i.e., OH groups + H\textsubscript{2}O\textsuperscript{mol}). We used the Beer-Lambert approximation (Morizet et al., 2002, 2007):

$$
C_i = \frac{Abs_i \cdot 18.02}{\rho \cdot d \cdot \epsilon} \tag{6}
$$

where $C_i$, the concentration of i-species, is a function of the integrated absorbance of the peak vibration ($Abs_i$) times the molecular mass of the water molecules (18.02, g.mol\textsuperscript{-1}) and divided by the product between the density ($\rho$, g.L\textsuperscript{-1}), the thickness of the sample ($d$, cm) and the integrated molar extinction coefficient for the considered specie ($\epsilon$, L.mol\textsuperscript{-1}.cm\textsuperscript{-2}).

Reliable quantification can only be achieved if suitable values for $\epsilon$ are available. In order to be consistent, we have used for all samples the $\epsilon$ values derived by Stolper (1982a) and
valid for basaltic composition: 200 L.mol\(^{-1}\).cm\(^{-2}\) for OH groups and 300 L.mol\(^{-1}\).cm\(^{-2}\) for H\(_2\)O\(^{mol}\). Vibration at 3550 cm\(^{-1}\) for total water was not used as the total peak could not be obtained, especially for high water content. A density value of 2537 g.L\(^{-1}\) was used and determined from Lange and Carmichael (1987) glass density model. The spectra for each sample were normalised to 100 \(\mu\)m thick.

The calculated H\(_2\)O\(^{m}\) is indicated in Figure 3. The value is based on the FTIR measurements of at least 3 different spectra. The error on each value is calculated from the standard deviation analysis of the sum OH, H\(_2\)O\(^{mol}\) concentrations. For KB3, the OH signal is very weak. The H\(_2\)O\(^{tot}\) error reported in the Figure 3 is probably underestimated and may be closer to +/- 0.1 wt.% rather than +/- 0.01 wt.%. 

4 Discussion:

4.1 Raman quantification factor (F, G) ratios for \(^{13}\)C species:

From the Placzek’s polarizability theory, a quantitative analysis of the species molar fraction can be achieved (Schrötter and Klöckner, 1979; Wopenka and Pasteris, 1986, 1987; Dubessy et al., 1989; Burke, 2001).

Applying Eq. (4) to our binary fluid system (CO\(_2\)-CO), the \(X(CO_2)\) can be calculated using the peak area (A) or height (H):

\[
X(CO_2) = \frac{A(CO_2)}{F(CO_2)} \frac{A(CO)}{F(CO)} \frac{A(CO)}{F(CO)} \tag{7}
\]
\[
X(\text{CO}_2) = \frac{\frac{H(\text{CO}_2)}{G(\text{CO}_2)}}{\frac{H(\text{CO}_2)}{G(\text{CO}_2)} + \frac{H(\text{CO})}{G(\text{CO})}}
\] (8)

F and G are the Raman quantification factors used with the area and the height of the peaks respectively. Both values are of interest and provide complementary information. G-factor value has been constrained for other binary gas mixtures. Seitz et al. (1993, 1996) derived the G- and F-factor for CH4-N2 and CH4-CO2 gas mixture. For example, the G-factor between CH4 and CO2 shows more variability compared to the F-factor as a function of pressure. Such difference for CH4-CO2 could be the result of modified molecular interactions upon pressure inducing a peak line broadening.

Eq. (7) and (8) are linear. Area and height should be measuring the same quantities. In Figure 4, we have reported the peaks areas \(A_i\) versus the peak heights \(H_i\) for all observed species and resonance (CO2 \(2\nu_2\), \(\nu_1\) and CO for \(^{13}\text{C}\) and \(^{12}\text{C}\)) which were measured from all inclusions by LRM. There is a clear linear trend \(A_i = 4.70 H_i\) relating the height and the area of a given resonance which is consistent with the linear aspect character of Eq. (7) and (8).

We consider a mixture between CO2 and CO in the fluid phase. We make the assumption that the small amount of CH4 does not intervene in the \(X(\text{CO}_2)\) calculation. The Raman scattering coefficient for CH4 is 8 times higher than for CO2 or CO (Seitz et al., 1996; Burke, 2001). In KB16, the area measured for the presence of CH4 is roughly equivalent to the area measured for CO. We estimated that CH4 would only represent less than 5\% \((X(\text{CH}_4) < 0.05)\) in the fluid phase which is probably within the error of the Raman measurements.
With CO₂ and CO, Eq. (4) and (5) can be rearranged and can be expressed as a function of the F- and G-factors:

\[
(1 - X(CO₂)) \times A(CO₂) = X(CO₂) \times A(CO) \times \frac{F(CO₂)}{F(CO)}
\]  

(9)

\[
(1 - X(CO₂)) \times H(CO₂) = X(CO₂) \times H(CO) \times \frac{G(CO₂)}{G(CO)}
\]  

(10)

We obtain a linear relation between A (or H) and the F (or G) ratio passing through zero. Eq. (9) and (10) can be applied to either C isotope but we first consider the ¹³C for which NMR data is available. A(CO₂) stands for the area of the CO₂ peak which is either the 2ν₂ or ν₁ resonance. We considered the resonance of the CO₂ diad individually, although, Dubessy et al. (1989) suggested using the sum of the peak areas for quantification. Seitz et al. (1996) on the other hand concluded that it was more reliable to use the 2ν₂ resonance instead of the peaks sum. We will see in fact that each resonance may be used without significant difficulties.

Application of Eq. (9) and (10) is represented in Figure 5. We use the values obtained from NMR as an input for the X(¹³CO₂NMR) and combined it with either the area of Raman peaks (Figure 5A and 5B) or the height of Raman peaks (Figure 5C and 5D). In order to be consistent with the NMR results, we only used the Raman information for the ¹³C isotope.

Although, we obtain a good linear correlation (dashed line) with 95% confidence interval (dotted lines) for the calibrations of the F ratios using the area of the peaks (Figure 5A and 5B) a slight variability may be observed. We do not preclude from compositional
heterogeneity in the fluid inclusions. Slight change in the fluid inclusion composition is only visible through Raman analysis but is averaged out with the NMR bulk method. The height of the peak gives a good correlation when using the $2\nu_2$ resonance (Figure 5C and 5D). We obtain an F($2\nu_2$ $^{13}$CO$_2$/F($^{13}$CO) = 1.956 ± 0.028. Calibration of the G-factor provides similar results with G($2\nu_2$ $^{13}$CO$_2$/F($^{13}$CO) = 1.809 ± 0.035. This is in agreement with the fact that the area and height of the peaks should be measuring the same quantity. Up to now, no data are available for the calibration of the $\nu_1$ CO$_2$ peak quantification factor. We find a value F($\nu_1$ $^{13}$CO$_2$/F($^{13}$CO) = 0.526 ± 0.014. Using the height of the peak, we calibrated the G ratio for $\nu_1$ resonance of $^{13}$CO$_2$. In Figure 5D, it seems there are multiple correlations. We can separate the dataset in 2 parts: the first represented by KB16 and the second represented by the other samples, KB3, 5 and 8. Two trends are identified. For KB16, the G factor is roughly 0.3 whereas for KB3, 5 and 8, the G factor is around 0.5. The main difference in between the samples is the $^{13}$C enrichment which is different. KB16 has roughly 80% in $^{13}$C isotopes. KB3, 5 and 8 are enriched at a lower level (~40%). The multiple trends aspect only concerns the $\nu_1$ calibration and this behaviour is not observed for the $2\nu_2$. The use of the $\nu_1$ resonance height may not be suitable for relevant calibration of randomly enriched samples. The shape of the $\nu_1$ resonance peak is not changing linearly with the $^{13}$C enrichment of the samples.

4.2 Raman quantification factor applied for $^{12}$C species:

Calibration results obtained for $^{13}$C-species are extended to $^{12}$C-species to test if whether or not CO$_2$ and CO molecules in both isotopes possess the same Raman cross-sections (i.e. F or G ratio for $2\nu_2$ $^{12}$CO$_2$/ $^{12}$CO are similar). In other words, we test the assumption
that $^{13}$C and $^{12}$C species do not fractionate in the fluid phase and enter the melt phase indifferently. We regard the reaction $\text{CO}_2 \leftrightarrow \text{CO} + \frac{1}{2} \text{O}_2$ as equivalent in between the isotopes on a thermodynamic point of view.

Using the $X(^{13}\text{CO}_2)$ measured from NMR, combining it with the integrated area and the height of the $2\nu_2$ $^{12}\text{CO}_2$ and $^{12}\text{CO}$, we have derived the F and G ratio for the $^{12}$C species which is represented in Figure 6A and 6B. We only used this approach for $2\nu_2$ resonance for clarity as we suggested $\nu_1$ not to be reliable enough for calibration as seen in Figure 5D. Although we take the $X(^{13}\text{CO}_2)$ from NMR, if a significant difference would exist in between the two isotopes then several trends will be expected. For example in Figure 5D, we observe 2 different trends for the calibration of the G factor of the $\nu_1$ resonance. The 2 trends are likely to be explained by the different enrichment in $^{13}$C isotope: KB16 is more enriched in $^{13}$C (~80%) than the other samples (~40%).

Values derived from the linear fit are 1.856 ± 0.057 and 1.756 ± 0.054 for F and G factor respectively for $^{12}$C isotope. Those values are in the same order as for the calibration obtained for $^{13}$C species. The errors on the values are larger than for $^{13}$C species G and F factors. The data points for KB3 in particular show a large scatter compared to the other samples. Thus, as KB3 is included in the fit, it represents the major source of error in the final values. Removing KB3 data points would probably increase the obtained value for F and G factors. A slight heterogeneity in the KB3 fluid inclusions compositions may be responsible for the large scatter.

The F ratio given by Wopenka and Pasteris (1987) for the $2\nu_2$ $^{12}\text{CO}_2$ and $^{12}\text{CO}$ is reported for comparison (1.55 corresponding to 1.52 and 0.98 for $2\nu_2$ $^{12}\text{CO}_2$ and $^{12}\text{CO}$ Raman scattering cross-section).
The F ratios value obtained from linear regression in Figure 6 ($2\nu_2^{13}\text{CO}_2$ vs. $^{13}\text{CO}$) is only slightly different from the one established by Wopenka and Pasteris (1987) determined for a RAMANOR U-1000 spectrometer. The difference in between the calibration values may be due to either different Raman efficiency coefficient value ($\zeta$) or a true effect of the fluid inclusions trapping conditions. Synthetic fluid inclusions in Wopenka and Pasteris (1987) were formed at less than 1.5 MPa. Although we did not measure the pressure inside the inclusions, it is likely that the pressure inside our fluid inclusions is higher. The different inclusion pressure could have a possible effect on the quantification factor.

The supposed change for F with pressure has implications for the quantification of high pressure fluid species through LRM. If one needs to determine the molar fraction of a gas species entrapped at high pressure then a proper calibration needs to be done for the quantification factor as the reported value for low pressure fluid inclusions may not be suitable for high pressure domain. Yet, it is currently unknown what behaviour the F and G factor could adopt at very high pressure. Using non suitable value for high pressure fluids may therefore induce a significant error on the determination of the gas mixture composition and thus on the molar fractions calculations.

Because we obtain similar value for the F and G factors for the 2 different isotopes, we conclude that our analytical approach (combining NMR and LRM analysis to constrain the Raman quantification factor) is adequate. Moreover, we also suggest that $^{12}\text{C}$ and $^{13}\text{C}$ species have comparable behaviour for Raman scattering. Thus, both isotopes can be quantified through LRM with the same F or G factor.
4.3 CO₂ molar fraction recalculated from LRM and correction for H₂O content in the fluid phase:

The calibration has been performed considering only CO₂ and CO are present in the fluid phase and neglecting the presence of CH₄ as C-species. However, CO₂ and CO species should be in thermodynamic equilibrium with H₂O in the fluid phase as H₂O is present in the melt. We estimated the H₂O molar fraction in the fluid phase (X(H₂O)) according to Eq. (11):

\[ X(H₂O) = \frac{f(H₂O)}{P_{tot} \cdot \phi(H₂O)} \]  

(11)

where \( f(H₂O) \) stands for the H₂O fugacity; \( P_{tot} \) is the total pressure of the experiment; \( \phi \) is the H₂O fugacity coefficient. The \( f(H₂O) \) was estimated from the model of Dixon et al. (1995) which relates the H₂O solubility in the melt to the H₂O fugacity in the fluid. H₂O fugacity coefficient is determined from the equation of state given in Shi and Saxena (1992). The results on the X(H₂O) are reported in Table 2. Values oscillates from almost 0 for KB3 and up to ~20% for KB12. Although most of the calculated values are below 10% for the other samples. The error on the X(H₂O) was calculated considering an error of +/-10 bars on the determination of the \( f(H₂O) \).

Using the calibration of G and F factors obtained for the 2ν₂ CO₂ resonance (see Figure 5 and 6), we have recalculated the X(¹³CO₂) and X(¹²CO₂) uncorrected for X(H₂O). Results are given in Table 2. Each value represents the average and the standard deviation for X(CO₂) obtained from all the fluid inclusions analyzed by LRM.

The X(CO₂) in the fluid phase has been recalculated accounting for the amount of H₂O present in the fluid phase. According to mass balance and considering ¹²C and ¹³C isotopes do not produce significant fractionation, we have:
\[
X(H_2O) + X(CO_2) + X(CO) = 1 \tag{12}
\]

Correction on the X(CO\textsubscript{2}) was done according to the following equation:

\[
X(CO_2)_{\text{corrected}} = \frac{1 - X(H_2O)}{\alpha + 1} \tag{13}
\]

where \( \alpha = X(CO)/X(CO_2) \) determined from either NMR or LRM prior correction.

The X(CO\textsubscript{2}) corrected value is also reported in Table 2. For a low X(H\textsubscript{2}O), the X(CO\textsubscript{2}) corrected value does not significantly change from the uncorrected one. In KB3, X(CO\textsubscript{2}) changes from 0.769 to 0.766 for uncorrected and corrected value respectively. It is not the case for KB12 for which the X(CO\textsubscript{2}) is decreased from \(~0.93\) to \(~0.75\) between uncorrected and corrected value.

In Figure 7, the corrected X(CO\textsubscript{2}) determined by LRM are plotted against the corrected X(\textsuperscript{13}CO\textsubscript{2}) measured by NMR. The 1:1 line is also reported for comparison. The plot is presented in the range of molar fraction of interest (0.5 to 0.9).

The obtained calibration is satisfactory for most of the samples as the X(CO\textsubscript{2}) LRM reproduces well the X(\textsuperscript{13}CO\textsubscript{2}) NMR. \textsuperscript{13}C species abundances are reproduced precisely by the produced calibration.

Reproducing the X(\textsuperscript{12}CO\textsubscript{2}) by LRM appears more difficult especially for KB16 (\(~0.68\) for X(\textsuperscript{12}CO\textsubscript{2}) by LRM versus \(~0.61\) by NMR). One source of disagreement can potentially be the different enrichment in \textsuperscript{13}C observed in KB16; however this point would have to be confirmed with further fluid inclusion analyses on different \textsuperscript{13}C/\textsuperscript{12}C ratios.

4.4 Oxygen fugacity calculation from H\textsubscript{2}O and CO\textsubscript{2} equilibrium reactions:
In the fluid phase, CO$_2$, CO and O$_2$ are in thermodynamic equilibrium according to Eq. (2). Using the thermodynamic formalism described by Shi and Saxena (1992), we have calculated the $f$(O$_2$) within each sample. The Eq. (2) equilibrium constant ($K$(CO$_2$)) is related the X(CO$_2$) and X(CO) measured in the fluid according to the relation:

$$K(CO_2) = \frac{X(CO)\phi(CO)\cdot f(O_2)^{\frac{1}{2}}}{X(CO_2)\phi(CO_2)}$$  \hspace{1cm} (14)$$

where $\phi$ is the fugacity coefficient determined from equation of state and $K$ from the thermodynamic database given in Shi and Saxena (1992).

The redox state of the fluid ($f$(O$_2$)) is calculated following two methods. The first one is based on the CO$_2$-CO-O$_2$ equilibrium from fluid inclusion data and the second one on the H$_2$O-H$_2$-O$_2$ equilibrium using $f$(H$_2$) known from experimental conditions and $f$(H$_2$O) derived from solubility of water in the melt at experimental P and T conditions. As stated before, H$_2$O$^m$ is in equilibrium with H$_2$O$^n$, which implies that a(H$_2$O)$^n$ = a(H$_2$O)$^m$ or $f$(H$_2$O)$^n$ = $f$(H$_2$O)$^m$. If the redox state ($f$(O$_2$)) is controlled by H$_2$ fugacity, according to Eq. (1), the relationship between different gas fugacity is the following:

$$K(H_2O) = \frac{f(H_2O)}{f(H_2)\cdot f(O_2)^{\frac{1}{2}}}$$  \hspace{1cm} (15)$$

where $f$ represents the fugacity of the different species. K(H$_2$O) is calculated from thermodynamic data in Shi and Saxena (1992). The $f$(H$_2$) are experimentally applied values listed in Table 1.
Results are given in Table 3 listing the log $f(O_2)$ calculated with the two different methods and plotted in Figure 8. We choose to take the average of the log $f(O_2)$ (column 6) as representative of the LRM fluid inclusion analyses. The standard deviation on the calculated log $f(O_2)$ values is also represented. We only used the X(CO$_2$)-X(CO) values corrected for X(H$_2$O) and shown in Table 2 to calculate the log $f(O_2)$. However, calculation with uncorrected X(CO$_2$)-X(CO) values changes the log $f(O_2)$ by only 0.01 log unit.

We observe in Figure 8 a large disagreement between the two calculation methods for KB3. This discrepancy may be explained by the remarkably low water content in KB3. H$_2$O$^m$ estimation by FTIR is therefore more difficult and subject to very large error. The solubility of H$_2$O in silicate melt is a non linear function of $f(H_2O)$. For 0.27 wt.% H$_2$O$^m$ we found a $f(H_2O) = 10$ bars but owing to the associated analytical error of H$_2$O$^m$ determination, we can not exclude this value to be 2 to 3 times higher which would change the log $f(O_2)$ by almost 1 unit.

Moreover such a value (log $f(O_2)$ ~-13 corresponds to extremely reducing conditions well below the graphite saturation. Those reducing conditions would induce a change in the appearance of the sample which should be totally blackened by the graphite crystals. The $^{13}$C-MAS NMR would also have revealed the presence of very reducing species such as SiC or even AlC groups. This is not the case in the spectra (NMR or LRM). The NMR results on the X(CO$_2$) cannot be challenged as it is a more quantitative method than the FTIR. We suggest the H$_2$O$^m$ method to calculate the $f(O_2)$ is less reliable for our glasses with low water content in regard of 1) the possible large error on the FTIR.
measurements at low $H_2O^m$ and 2) the corresponding large uncertainty on the fugacity estimate.

Another possibility is the departure from 1:1 line. It appears there is also an offset of 0.5 to 1.0 log unit for KB5 and KB8 respectively. Either it represents the absolute error on the $\log f(O_2)$ calculation or one of the two method is not calculating the real $\log f(O_2)$ at equilibrium.

For example, compositional changes can occur during quenching either in the fluid phase or the melt phase (Zhang et al., 1995; Morizet et al., 2001). Thermodynamic calculations are performed at run temperature, however the fluid inclusion spectroscopic measurements are conducted at room temperature. Therefore, the $X(CO_2)$ measured may not be entirely representative of the high temperature fluid composition. This hypothesis could only be tested with in-situ measurements.

On the other hand, for KB16, 10 and 12 we obtain an excellent agreement in between the two methods. KB10 and 12 were not included into the LRM calibration but were used as an independent test. The fact that KB10 and KB12 show an excellent correlation is a very strong witness for the reliability of the Raman quantification factor (F and G) calibration presented in this work.

5 Summary:

Determining the fluid inclusion composition relies on micro-analytical techniques such as LRM. LRM requires a relevant calibration for measuring the gas molar fraction within the fluid inclusions.
We have combined LRM and $^{13}$C-MAS NMR analyses to produce a calibration for CO$_2$-CO gas mixture entrapped in synthetic fluid inclusions quenched from high temperature (1200-1250°C) and high pressure (2-3 kbars) experiments. The applied method with the use of NMR measurement has the major advantage to be independent of fluid density and is only function of fluid composition.

First, the calibration was applied to $^{13}$C species. We have determined the F and G factors for $^{13}$CO$_2$ and $^{13}$CO. From the CO$_2$ Fermi diad, we obtain a well defined unique linear trend when using the $2\nu_2$ $^{13}$CO$_2$ resonance whereas the use of the $\nu_1$ resonance produce strange behaviour. We suggest that only the use of the $2\nu_2$ CO$_2$ resonance is suitable for determining the fluid composition in CO$_2$-CO mixture.

Second, considering $^{12}$C and $^{13}$C isotopes behave similarly, we have extended our calibration procedure with $^{12}$CO$_2$-$^{12}$CO species identified in the fluid inclusion and in equilibrium with $^{13}$C species. We found values for F and G factors which are close to $^{13}$C species F and G values. It implies that 1) $^{12}$C and $^{13}$C species are not strongly fractionated in the fluid phase and may be considered almost equally on a thermodynamic point of view, 2) Although the chosen approach is likely to be more robust for $^{13}$C than for $^{12}$C species, our calibration may be applied to both isotopes indifferently.

The observed values for F and G are different from previously published value for F and G. The difference may be the result of the 1) different entrapment pressure or 2) change in the Raman efficiency coefficient. Most of the reported fluid composition calibrations have dealt with low pressure synthetic inclusions. Up to now, no data for F and G factors are available for high pressure domain and this point needs to be investigated further.
X(CO₂) has been recalculated according to the calibration results, it was then used to calculate the log f(O₂) in the fluid phase. The calibration has also been tested using 2 independent samples which were not used in the calibration process. There is an excellent agreement between the log f(O₂) calculated for those samples from fluid inclusions analyses and from water content within the melt.

A reasonable agreement is observed between the log f(O₂) calculated from 1) the fluid inclusions analyses and 2) H₂O dissolved in the melt. However, at low H₂O content, there is a strong departure of the log f(O₂) from the 1:1 line suggesting that log f(O₂) calculation method from H₂O dissolved in the melt may not be suitable under these conditions. Thus, fluid inclusions analyses may be considered as an alternative approach for determining the log f(O₂) in dry or nearly dry conditions.

Alternatively, a change in the fluid phase composition during quenching could explain this disagreement. However, only in-situ measurements of the fluid phase composition may be able to test this hypothesis.

Acknowledgements:
The authors are grateful to the University of Orléans, University of Nantes, the CNRS and INSU for their financial support and access to analytical facilities. We also thank Dr. R. Champallier and Mr. E. Lemenn for their technical help. We also would like to thank Prof. J. Dubessy and an anonymous reviewer for the constructive and helpful reviews.
Reference list:


Kerkhof, A.M. van den, 1988a. Phase transitions and molar volumes of CO$_2$-CH$_4$-N$_2$ inclusions. Bulletin de Minéralogie 111, 257-266.


Figure caption:

Figure 1: Fluid inclusions pictures taken on different samples. The picture is taken through the objective x60 and the scale is represented (25 μ).
Figure 2A: $^{13}$C-MAS NMR spectra for KB3 and 5. KB3 spectra were acquired with different recycle delay ($D_1 = 2$ and 20 sec.). Several species are identified: 125, CO$_2^{\text{fl}}$; 180, CO$_2^{\text{fl}}$; 160-175 ppm, CO$_3^{2-\text{melt}}$. The $X^{(13\text{CO}_2)}$ for each is added and was calculated from the ratio of the integrated area for fluid species.
Figure 2B: Fluid inclusion Raman spectra showing the signature of the CO₂-CO gas mixture for $^{12}$C and $^{13}$C isotopes. The CO₂ Fermi is represented by 2 main vibrations: $2\nu_2$ (1385, 1360 cm$^{-1}$ for $^{12}$CO₂ and $^{13}$CO₂) and $\nu_1$ (1285, 1260 cm$^{-1}$ for $^{12}$CO₂ and $^{13}$CO₂);
and accompanied by a “Hot band” vibration at 1405 cm\(^{-1}\). CO shows 2 bands at 2140 and 2090 cm\(^{-1}\) for \(^{12}\)C and \(^{13}\)C respectively.

Figure 3: FTIR spectra showing the water vibrations: OH groups, 4500 cm\(^{-1}\); H\(_2\)O\(^{\text{mol}}\), 5200 cm\(^{-1}\). The dashed line represents the baseline placed prior to integration of the vibration peaks. The water content (H\(_2\)O\(^{\text{m}}\)) is also indicated and was derived from Beer-Lambert approximation (see section 3.2.3 for detail).
Figure 4: LRM peak area (A) versus peak height (H) for the observed gas resonances (2ν₂ CO₂, square; ν₁ CO₂, circle; and CO, triangle) and for both isotopes (¹²C, filled symbol; ¹³C, open symbol).
$G(2v_2^{13}CO_2) / G(^{13}CO) = 1.809 \pm 0.035$
Figure 5: Raman quantification factor (F and G) calibration for $^{13}$C isotopes. A, calibration for $F(2\nu_2 \text{CO}_2) / F(\text{CO})$; B, calibration for $F(\nu_1 \text{CO}_2) / F(\text{CO})$; C, calibration for $G(2\nu_2 \text{CO}_2) / G(\text{CO})$; D, calibration for $G(\nu_1 \text{CO}_2) / G(\text{CO})$. 
(1 - $X^{13}CO_2$). Area $^{12}CO_2 2v$

$X^{13}CO_2^{NMR}$. Area $^{12}CO$

$W. & P. (1987) = 1.55$

$F(2v_2^{12}CO_2) / F(12CO) = 1.856 \pm 0.057$
Figure 6: Raman quantification factor (F and G) calibration for $^{12}$C isotopes. A, calibration for $F(2\nu_2 \text{CO}_2) / F(\text{CO})$; B, calibration for $G(2\nu_2 \text{CO}_2) / G(\text{CO})$. F factor value for $^{12}\text{CO}_2$ and $^{12}\text{CO}$ from Wopenka and Pasteris (1987) is also reported.
Figure 7: Recalculated $X(\text{CO}_2)$ from $2\nu_2$ CO$_2$ LRM calibration and plotted against $X(^{13}\text{CO}_2)$ measured from NMR. The circles and squares represent the $X(\text{CO}_2)$ for $^{13}$C and $^{12}$C respectively. Open symbols are obtained from G-factor and filled symbols from F-factor.
Figure 8: Log \( f(O_2) \) calculated from \( X(CO_2) \) versus log \( f(O_2) \) calculated from \( H_2O \) dissolved in the glass. The log \( f(O_2) \) is calculated from \( X^{13}CO_2 \) NMR (circle) and the average values from LRM calibrations (square). NMR value is only available for KB16, 3, 5 and 8.
### Tables:

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<th>Sample</th>
<th>P_{\text{tot}} (MPa)</th>
<th>Temperature (°C)</th>
<th>P(H_{2}) (bars)</th>
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Table 1: IHPV synthesis conditions for COH fluid bearing glasses. P_{\text{tot}} represents the final pressure during the experiment. The P(H_{2}) is the partial pressure of H_{2} at the experimental conditions. The H_{2}O^m was determined by FTIR analysis (see section 3.2.3 and Figure 2 for details). The error is calculated from standard deviation analysis. The X(^{13}\text{CO}_2) was determined by peak integration of the ^{13}\text{C}-\text{MAS NMR} spectra. Value are uncorrected for the presence of H_{2}O in the fluid phase. ^# NMR spectra were acquired in different conditions. For KB3, two different values for D1 were used: 2 and 20 sec.. KB16 NMR spectra were acquired with 2 different pulse sequences: SPE and spin-echo (see section 2.3 for details).
Table 2: Calculated species molar fractions in the fluid phase. X(\text{H}_2\text{O})^a is derived from Dixon et al. (1995) relating total water content in the melt to fluid phase composition. X(\text{CO}_2) values are corrected for water content in the fluid phase (see section 4.3 for details). Error bars on the corrected values are based on error propagation from standard deviation. Error on the uncorrected values is based on the standard deviation from the fluid inclusion analyses by LRM. X(\text{CO}_2) is determined by LRM using both the calibration derived from peak integrated area (from F) and the calibration from peak height (from G). For clarity, X(\text{CO}) corrected values are not indicated but can be determined according to mass balance (Eq. (12)).

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<th>Sample</th>
<th>X(\text{H}_2\text{O})^a</th>
<th>X(\text{\textsuperscript{13}CO}_2) \text{NMR}</th>
<th>X(\text{\textsuperscript{13}CO}_2) \text{LRM}</th>
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<td>0.661 ±0.015</td>
<td>0.614 ±0.006</td>
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\(^a\) X(\text{H}_2\text{O}) is derived from Dixon et al. (1995) relating total water content in the melt to fluid phase composition. \(^b\) X(\text{CO}_2) values are corrected for water content in the fluid phase (see section 4.3 for details). Error bars on the corrected values are based on error propagation from standard deviation. \(^c\) Error on the uncorrected values is based on the standard deviation from the fluid inclusion analyses by LRM. \(^d\) X(\text{CO}_2) is determined by LRM using both the calibration derived from peak integrated area (from F) and the calibration from peak height (from G). For clarity, X(\text{CO}) corrected values are not indicated but can be determined according to mass balance (Eq. (12)).
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<th>Sample</th>
<th>Log f(O₂) from H₂O&lt;sup&gt;m&lt;/sup&gt;</th>
<th>Log f(O₂) from X(CO₂) NMR</th>
<th>Log f(O₂) from &lt;sup&gt;13&lt;/sup&gt;CO₂ LRM</th>
<th>Log f(O₂) from &lt;sup&gt;12&lt;/sup&gt;CO₂ LRM</th>
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Table 3: Recalculated X(CO₂) from LRM fluid inclusions analyses: for <sup>13</sup>C and <sup>12</sup>C and from F and G calibration. The log f(O₂) is also represented and calculated using the MRK equation from Shi and Saxena (1992) from X(CO₂) or the H₂O<sup>m</sup> in the glass.
## Appendix: Data for fluid inclusion analyses by LRM

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